

Sample 50µl from MeOH supernatant of feeding
page (total 4 ml volume)

$$\begin{array}{l} 50\mu\text{l} : 38,406 \\ \underline{50\mu\text{l}} : 38,886 \end{array} \left. \vphantom{\begin{array}{l} 50\mu\text{l} \\ \underline{50\mu\text{l}} \end{array}} \right\} 38,646 \text{ counts}/4 \text{ min} = 9661.5 \text{ cpm}$$

$$\frac{9661 \text{ cpm}}{0.05 \text{ ml}} = 1.932 \times 10^5 \text{ cpm/ml}$$

⇒ 7.7×10^5 cpm in total solubilized sample

by P.24 — there should be 2.2×10^6 counts in 10 mg!

$$\begin{array}{l} 7.7 \times 10^5 \\ 5.5 \times 10^4 \end{array} \text{ counts} = 14 \text{ mg}$$

∴ I have $\frac{1}{4}$ the amount of product
that I want.

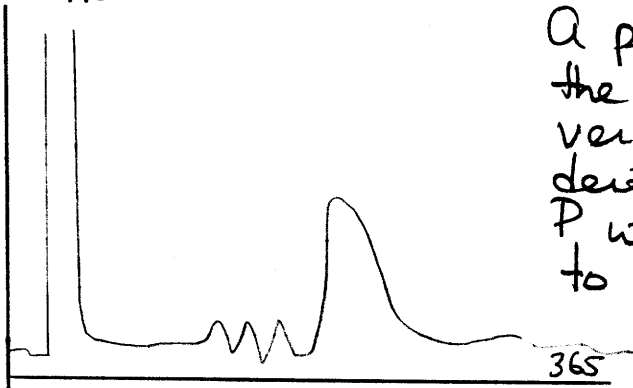
9-23-76 Further Processing of 8-25-2 (HCOOH-hydrolyzed DNA)

Sample 8-25-2 was prepared from 10 mg of adducted DNA - it should contain up to 30 μ g of adduct - I need adduct for identification studies, so this sample will be processed further.

Procedure

- sample is in a small RB - it is a whitish residue
- add 2 ml redistilled MeOH -
- add small stirring bar - stir for 1.25 hr. Observation - the sample only slowly went into suspension. I'm hoping to slowly dissolve the adduct, or a portion of it.
- remove the 2 ml
- add fresh 2 ml MeOH and spin for 1/2 hour.
- combine MeOH suspensions in polycarbonate tube
- centrifuge 12K x 10 min
- Count 50 μ l and inject 50 μ l into HPLC

Results:



A peak appeared at $\sim t_R$ of the adduct, but it was very broad - column problems developed. ~~As~~ explicitly, the P went \uparrow and it took (3) days to repair the damage